

0040-4039(94)01671-2

Taxol Chemistry. 7-O-Triflates as Precursors to Olefins and Cyclopropanes

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Abstract: 7-O-Triflates of baccatin III or of taxol analogs are convenient precursors in alternate syntheses of $\Delta^{6,7}$ -taxols and 7 β ,8 β -methano (cyclopropyl) taxols, two of the products initially obtained from reaction of taxols with methylDAST; hydrazine is an effective reagent for conversion of the 10-acetate group to the hydroxyl group.

Taxol, in addition to its exciting potential as an antitumor agent, presents a fascinating array of challenges to the organic chemist.¹ The highly functionalized diterpenoid nucleus of taxol not only provides a synthetic challenge but is subject to unusual and often unexpected chemical transformations. Contraction of rings A and B,² oxetane opening,^{2,3} epimerization at C-7,⁴ dienone formation,⁵ and cyclo-propane formation⁶ are some of the fascinating modifications of taxol which have been reported to date.

Independently of the work reported recently by Chen and coworkers,^{6a,6b} we have studied the reaction of a 2'-protected taxol with the fluorinating reagent, dimethylaminosulfur trifluoride (methylDAST). Our reaction of 2'-troc-taxol⁷ (1) with methylDAST⁶ gave 2'-troc-7-deoxy-7 α -fluoro-taxol (2, 33%) and 2'-troc-7-deoxy-7 β ,8 β -methano-taxol (3, 23%),⁹ results very similar to those reported by Chen and coworkers. However, we have isolated a third product from this reaction which is the olefin, 2'-troc-7-deoxy- $\Delta^{6,7}$ -taxol (4, 4%). Removal of the 2'-troc group gives 7-deoxy- $\Delta^{6,7}$ -taxol (5). The assignment of olefinic structures to 4 and 5 is based on three coupled, downfield signals for the C-5, C-6, and C-7 protons in the ¹H NMR spectra¹⁰ and was confirmed as described below. Elimination with formation of olefinis is a well documented side reaction in the reaction of alcohols with DAST.¹¹

Olefins are also produced in reactions of other taxol analogs with methylDAST. For example, the reaction with the 2'-troc derivative 7 of analog 6, in which the N-benzoyl group is replaced by benzyloxycarbonyl (Cbz), gave fluoride 8 (45%), cyclopropane 9 (18%), and olefin 10 (5%). Likewise, the reaction with the 2'-troc derivative 12 of analog 11, in which the N-benzoyl group is replaced by *t*-butyloxycarbonyl (BOC), gave fluoride 13 (47%), cyclopropane 14 (21%), and olefin 15 (5%). The isolation of olefins from these reactions is noteworthy because in every case the olefinic product is more potent than the alcohol precursor in assays which measure potential antitumor activity.¹²

The low yields of olefins produced by the methylDAST reactions called for development of an alternate source. For this, we found it possible to prepare and purify 7-O-triflates of either baccatin III (16) or of the taxol analog. The reaction of 16 with triflic anhydride in $CH_2Cl_2/pyridine gives baccatin III-7-OTf (17)$ in 80% yield.¹³ When 17 is warmed in solution in the presence of DBU, elimination of the 7-O-triflate proceeds smoothly to generate the 6,7-olefin 18 in 70% yield.¹⁴ The 6,7-olefins exhibit slightly enhanced sensitivity to acid catalyzed opening of the oxetane ring. Whereas the 2'-troc group of 4 can be



removed with zinc in HOAc-MeOH without rearrangement of the oxetane, 18 upon warming with acetic acid in $ClCH_2CH_2Cl$ rearranges to 19 (70%). The structures of both 18 and 19 have been confirmed by x-ray crystallographic studies.¹⁵



The $\Delta^{6.7}$ -analog 20, obtained by removal of the 2'-troc group from 15, now can be prepared by the alternate route shown in Scheme 1 which we have developed independently of that by Didier and coworkers.¹⁶ First, baccatin III-7-O-Tf (17) is coupled with (4S,5R)-N-Cbz-2-(2,4-dimethoxyphenyl)-4-phenyl-5-oxazolidinecarboxylic acid (21)^{9,16} to give 22 (87%, two epimers) from which the aminal groups are removed by exchange with acidic methanol to give 23 (93%). The Cbz group is removed from 23 by hydrogenolysis, giving the versatile intermediate 24 which is useful for attachment of a variety of N-substituents. Reaction of 24 with di-*t*-butylcarbonate provides the N-BOC compound 25 (77% from 23). Warming of 25 with DBU in THF generates the analog 20^{12,17} in 82% yield. Reaction of 24 with *t*-butylisocyanate provides the N-*t*-butylurea (TBU) compound 26 (74%). Warming of 26 with DBU in THF generates the analog 27^{12,18} in 75% yield.

The 7-O-triflates such as 17 also serve as excellent precursors for the 7 β ,8 β -methano (cyclopropyl) taxol analogs. Simply stirring a solution of 17 with silica gel (40-63 μ m, 1:60 w/w) in 1,2-dichloroethane at 60°C produces 7-deoxy-7 β ,8 β -methanobaccatin III (28)¹⁹ in 75% yield. When 28 is carried through a

Scheme 1



sequence of reactions analogous to those described in the preceding paragraph, we obtain 29 (the analog derived from 14 upon removal of the 2'-troc group). The influence of the C-9 substituent upon the reactivity of a 7-O-triflate is clearly seen by comparison of our results with those of Klein and coworkers.^{6c} Their report describes migration of the C-8 methyl to C-7 when a baccatin having both 7β - and 9α -hydroxyl groups is treated with triflic anhydride. The formation of an intermediate carbocation at C-8 is postulated, which undergoes further rearrangement by contraction of ring B from eight to seven carbons in size. Our results show that when there is a carbonyl group at C-9, the 7-O-triflates are relatively stable compounds which are converted to 6,7-olefins or 7,8-cyclopropanes under the conditions described above. Clearly, the electron withdrawing effect of the C-9 carbonyl group stabilizes the 7-O-triflates and completely inhibits carbocation formation at C-8.



As a final point of interest, we report that hydrazine (98%) in 95% ethanol is an excellent reagent for cleavage of the 10-acetate group when there is a free hydroxyl group at C-7. For example, 10-acetyltaxotere (11) is converted to taxotere²⁰ (30) in 86% yield at room temperature under these conditions. When the C-7 hydroxyl group is absent or masked by a protecting group, the removal of the 10-acetate by these conditions is much less effective.

References and Notes

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- 9. Upjohn PCT application, WO 9413655, June 23, 1994.
- 10. Spectral data for 5: ¹H NMR (CDCl₃, TMS) δ 8.17 (d, 2H), 7.75 (d, 2H), 7.64 (t, 1H), 7.36-7.55 (m, 10H, aromatic protons), 7.02 (d, 1H, -NH-), 6.21 (t, 1H, H₁₃), 6.20 (s, 1H, H₁₀), 6.06 (dd, 1H, H_{6 or 7}), 5.87 (d, 1H, H₃), 5.83 (m, 2H, H₂ and H_{6 or 7}), 5.10 (d, 1H, H₃), 4.79 (d, 1H, H₂), 4.44 (d, 1H, H₂₀₀), 4.32 (d, 1H, H₂₀₀), 4.00 (d, 1H, H₃), 2.39 (s, 3H, -CH₃), 2.23 (s, 3H, -CH₃), 1.87 (s, 3H, -CH₃), 1.70 (s, 3H, CH₃), 1.24 (s, 3H, -CH₃), 1.16 (s, 3H, -CH₃); mass spectrum: 836.3288, C₄₇H₄₉NO₁₃ + H requires 836.3282.
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- 12. The mouse L1210 leukemic cell assay [Cf., Li, L.H.; et.al., Cancer Res. 1992, 52, 4904] gives IC_{s0} values (μ M) of 0.017 for taxol, 0.0042 for 5, >0.1 for 7, 0.062 for 10, 0.0035 for 11, 0.0016 for 20, 0.013 for 31, and 0.0032 for 27. These results are from single assays. We thank Joan Culp for these assay results.
- Reaction conditions: Baccatin III (1.0 g) in CH₂Cl₂ (4.0 mL) and pyridine (3.4 mL) was cooled (-30 °C); Tf₂O (1.2 g) was added and the solution allowed to warm to RT. After 4 hrs, workup and chromatography (flash SiO₂, 5% CH₃CN/CH₂Cl₂) gave 17 (0.978 g).
- 14. Reaction conditions: 17 (0.97 g) and DBU (1.03 g) in THF (6 mL) were heated (reflux) for 4 hours.
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- 17. Spectral data for **20**: ¹H NMR (CDCl₃, TMS) δ 8.15 (d, 2H, J = 7.2 Hz), 7.61 (t, 1H), 7.51 (t, 2H, J = 7.5 Hz), 7.30-7.43 (m, 5H), 6.22 (s, 1H, H₁₀), 6.21 (t, 1H, H₁₃), 6.06 (dd, 1H, J = 5.6, 9.9 Hz, H₆), 5.87 (d, 1H, J = 9.6 Hz, H₇), 5.84 (d, 1H, H₂), 5.39 (d, 1H, J = 9.6 Hz, -NH- or H₃), 5.26 (d, 1H, H₃. or -NH-), 5.10 (d, 1H, J = 5.6 Hz, H₅), 4.61 (m, 1H, H₂.), 4.43 (d, 1H, J = 8.1 Hz, H₂₀), 4.30 (d, 1H, J = 8.2 Hz, H₂₀₀), 4.01 (d, 1H, J = 6.5 Hz, H₃), 2.39 (s, 3H, -CH₃), 2.33 (m, 1H), 2.24 (s, 3H, -CH₃), 1.86 (s, 3H, -CH₃), 1.76 (s, 3H, -CH₃), 1.34 (s, 9H, Me₃C-), 1.27 (s, 3H, -CH₃), 1.16 (s, 3H, -CH₃); mass spectrum, found: 832.3554, C₄₅H₅₃NO₁₄ + H requires 832.3544, 776, 551, 491, 369, 327, 105, 57 m/z.
- 18. Spectral data for 27: ¹H NMR (CDCl₃, TMS) δ 8.14 (d, 2H), 7.63 (m, 1H), 7.51 (m, 2H), 7.33 (m, 5H), 6.22 (s, 1H), 6.17 (m, 1H), 6.04 (m, 1H), 5.85 (m, 2H), 5.33 (m, 1H), 5.15 (m, 1H), 5.10 (d, 1H), 4.60 (m, 2H), 4.42 (d, 1H), 4.30 (d, 1H), 3.99 (d, 1H), 2.42 (s, 3H), 2.32 (m, 2H), 2.23 (s, 3H), 1.86 (s, 3H), 1.75 (s, 3H), 1.23 (m); mass spectrum: 831.3742; C₄₅H₅₄N₂O₁₃ + H requires 831.3704.
- 19. Spectral data for **28**: ¹H NMR (CDCl₃, TMS) δ 8.13 (dd, 2H, Ar), 7.61 (tt, 1H, Ar), 7.49 (t, 2H, Ar), 6.34 (s, 1H, H₁₀), 5.63 (d, 1H, J = 7.6 Hz, H₂), 4.83 (m, 1H, H₁₃), 4.75 (d, 1H, J = 0.9 Hz, H₃), 4.30 (d, 1H, J = 8.5 Hz, H₂₀), 4.18 (d, 1H, J = 7.6 Hz, H₃), 4.04 (d, 1H, J = 8.5 Hz, H₂₀), 2.48 (dt, 1H, J = 16.0 Hz, H₆), 2.39-2.20, (m, 2H, H_{14s,14b}), 2.26 (s, 3H, CH₃), 2.08 (s, 3H, CH₃), 2.03 (s, 3H, CH₃), 1.64 (m, 1H, H_{19b}), 1.35 (m, 1H, H₇), 1.23 (s, 3H, CH₃), 1.10 (s, 3H, CH₃); FAB mass spectrum: 569.2389, C₃₁H₃₆O₁₀ + H requires 569.2387.
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(Received in USA 19 July 1994; revised 23 August 1994; accepted 29 August 1994)